





Příprava studijních materiálů studijního programu "Veterinární virologie"

Advanced techniques in the diagnostics

of viral infections in science and research

Doktorský studijní program Veterinární virologie

Fakulta veterinárního lékařství

Veterinární univerzita Brno

Vytvoření doktorského studijního programu "Veterinární virologie" na Veterinární univerzitě Brno

Specifický cíl B: Tvorba nových studijních programů v progresivních oborech

Projekt NPO registrační číslo NPO_VETUNI_MSMT-16594/2022

Výstup č. 2, vazba na cíl projektu č. 2, volitelný indikátor U3



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Advanced techniques in the diagnostics of viral infections in science and research

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Compliance with general rules - biological risks Use of protective equipment - gloves, shield... Decontamination of the material with 10 % bly Appropriate waste management Glove!!!! Essential when manipulating RNA !











Full blood

Bone marrow

Serum/plasma

Smears from the mucous membrane

Cell cultures

Blood stains

Body fluids CSF Bronchoalveolar lavage Amniotic fluid Semen **Tissue samples** fresh / frozen Embedded in paraffin Hair









Anti - coagulant substances

EDTA

Sodium citrate

Heparin

PCR Inhibitors







Sample packing and shipping:

Freezing???

Avoid temperature changes

Avoid temperature extremes

The three-pack rule

- 1. Mechanically resistant packaging
- 2. Absorbent layer
- 3. Impermeable layer









Sample storage — DNA Blood, bone marrow, body fluids...

- 22-25 °C Unsuitable (<24 h)
- 2-8 °C Max. 72 hours
- 20 °C Not suitable
- 70 °C Depending on the application

Do not freeze blood and bone marrow before lysis of erythrocytes







Specimen storage — RNA Blood, bone marrow, body fluids...

- 22-25 °C: Unsuitable, up to 2 hours
- 2-8 °C: Unsuitable, up to 2 hours
- 20 °C: 2-4 weeks
- 70 °C: Suitable

Do not freeze blood and bone marrow before lysis of erythrocytes







DNA storage

< 4 months 1-3 years < 7 years > 7 years

 \longrightarrow \longleftarrow \longleftarrow

2–25 °C	2–8 °C	-20 °C	-70 °C

Not very suitable

Long term storage in ethanol







NK preparation - what to take into account??

The quantity or volume of DNA or RNA needed for test Number of samples

- Centrifuge capacity
- The speed of the methods used

Automated systems?

Microtiter plates

semi - automated systems









Precautions for working with RNA

RNA is a very unstable molecule !

Easily degraded

Sterile, plastic utensils marked "For **RNA** Use Only "

Always use gloves!

Specially treated water (DEPC)







With placing of NK

DNA in TE buffer at 4 °C for weeks at –20 °C to -80 °C (long term)

RNA in Rnase free, ultra pure water at -70 °C







Analysis and characterization of nucleic acids









NA analysis

DNA

- Amplification methods (PCR, LCR)
- Hybridization methods (Southern blot)
- Cleavage with restriction enzymes

RNA

- Amplification methods (RT-PCR)
- Hybridization methods (Northern blot)

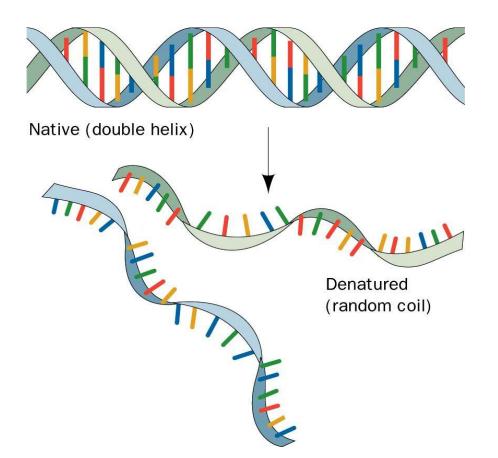








DNA denaturation

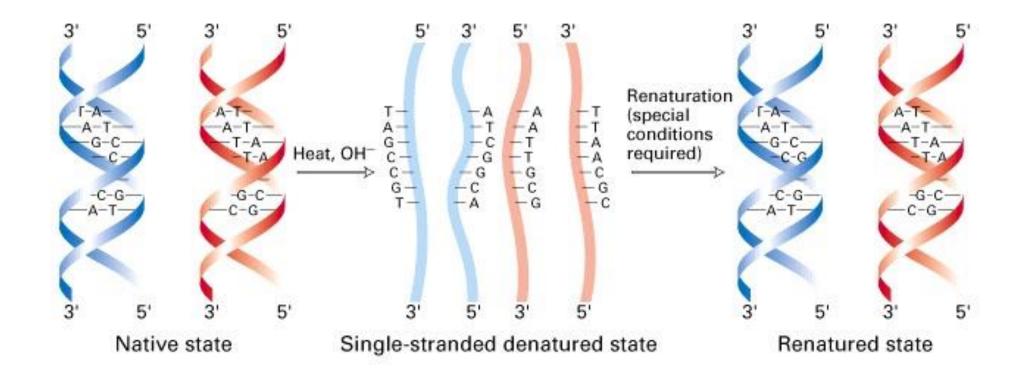




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Denaturation and renaturation





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NA amplification







Polymerase Chain Reaction

Enzymatic amplification of a specific DNA fragment



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Kary Mullis, PCR and the Nobel Prize

Knew that you could boil dsDNA to get ssDNA

Knew that you could use primers to initiate DNA synthesis

Knew that a cheap, commercial enzyme was available (Klenow fragment of *E. coli* DNA polymerase)







Kary Mullis, PCR and the Nobel prize

- He wanted to get a large amount of DNA from a single copy
- He originally used the "3 students " method
 - 1. Denaturing one
 - 2. Annealing one
 - 3. Extending one









Three steps of PCR

- **Denaturation template**
 - Usually 95 ° C
- Annealing primers
 - annealing temperature depends on the G+C content
 - High temperatures lead to high selectivity
- Extension of the new strand

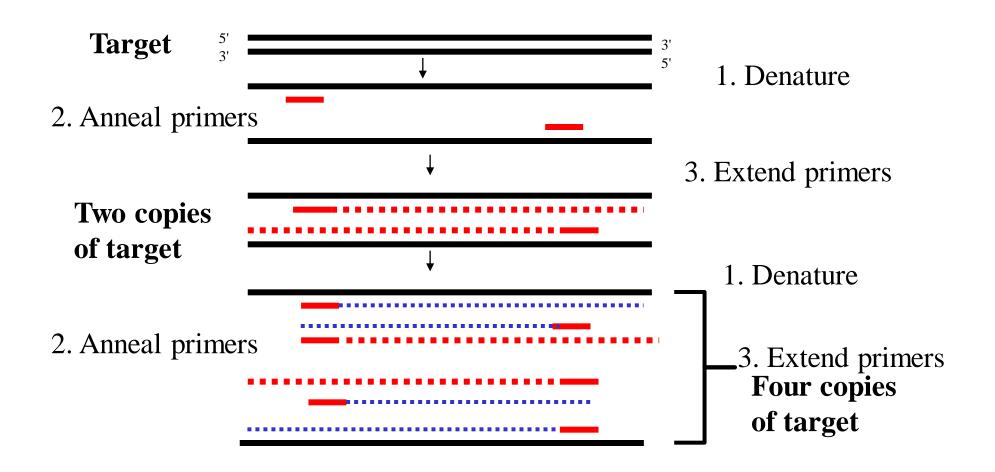








PCR AMPLIFICATION



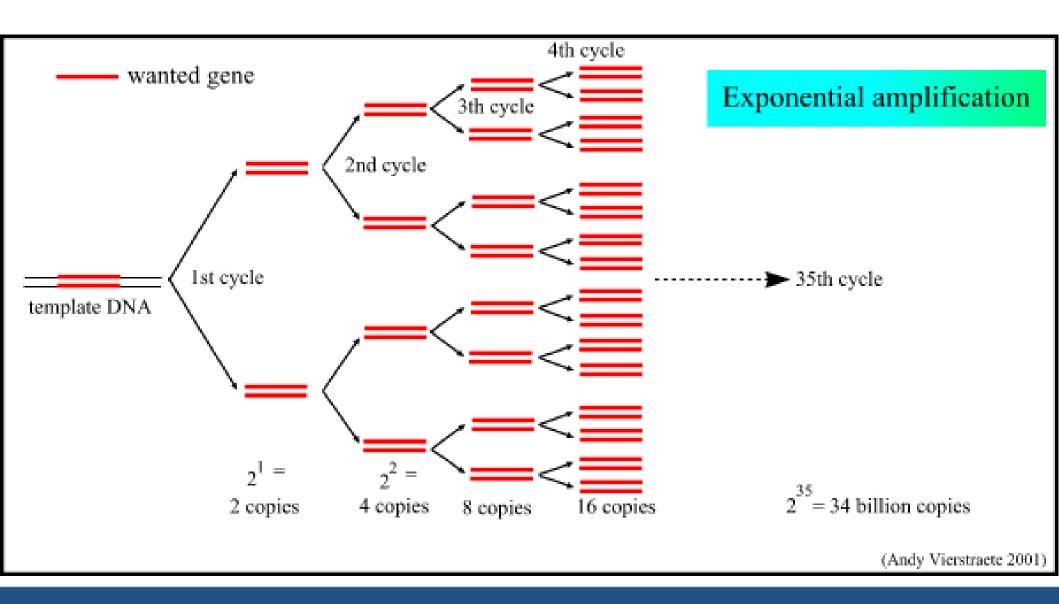
Národní

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PCR Primers

Primers are ssDNA fragments 18–30 bp long complementary to the sequences flanking the amplified region of the genome

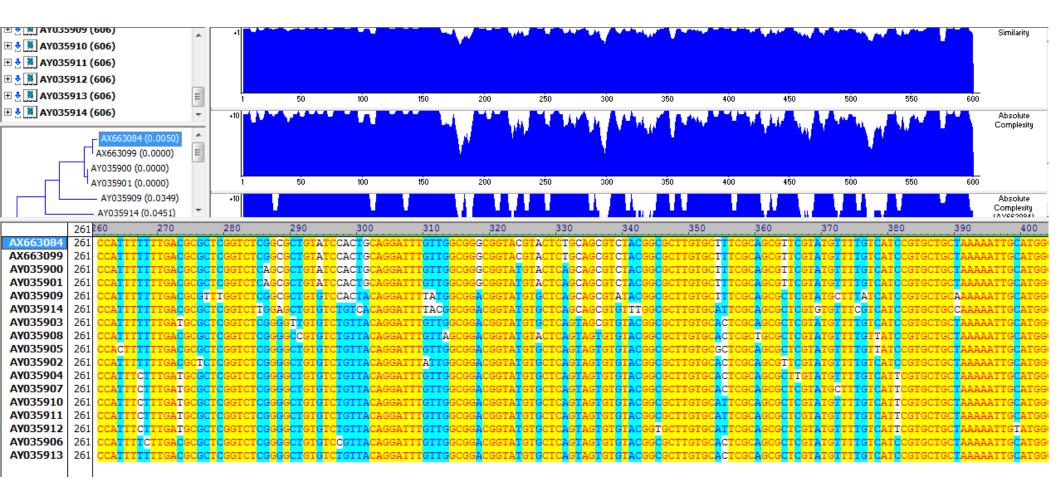
The primers are responsible for PCR specificity

The size of the PCR product depends on the primers position





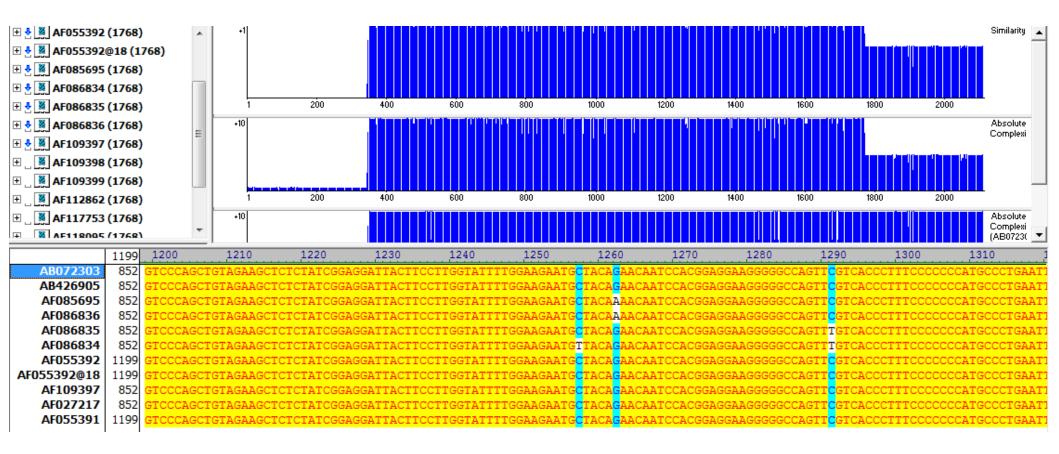




















PCR Master Mix standard

0.25 mM of each of the

0.2 mM of each of dATP, dCTP, dGTP, dTTP

50 mM KCl

- 10 mM Tris, pH 8.4
- 1.5 mM MgCl₂
- 2.5 U Taq Polymerase
- 10² 10⁵ template DNA
- 50 µl reaction volume









In Vitro amplification

How many amplicons will we get?

Number of amplicons = $A * 2^{n-2}$

n = number of PCR cycles

A = number of copies inserted into the reaction

Depletion of reaction components and polymerase errors lead to a plateau effect









Thermostable polymerases

Taq: Thermus aquaticus

Sequenase: T. aquaticus YT - 1

Restorase (Taq + *repair* enzyme)

Tfl: T. flavus

- Tth: T. thermophilus HB-8
- Tli: Thermococcus litoralis

Carboysothermus hydrenoformans (RT-PCR)

P. kodakaraensis (Thermococcus) (fast synthesis)

Pfu: Pyrococcus furiosus (accuracy)







PCR controls

Blank

- Contamination control
- All reagents except DNA

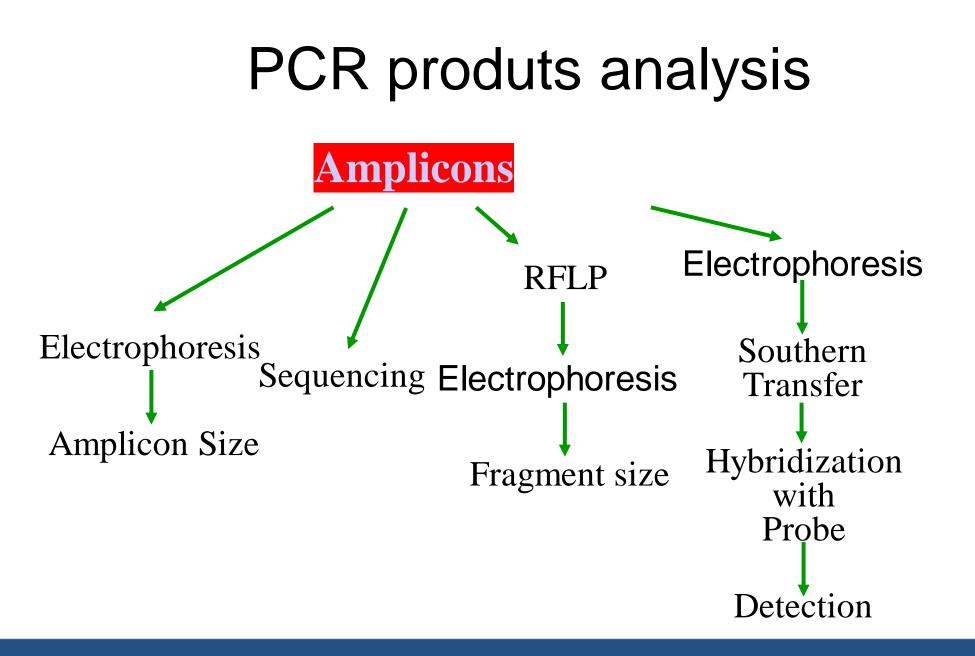
Negative control

- Specificity check
- All ingredients and DNA without target sequence **Positive control**
 - Sensitivity Control
 - All ingredients and known positive DNA

















Contamination of PCR reaction

Any DNA molecule containing the target sequence is a potential source of contamination

- The products of the preceding PCR reaction pose the greatest risk
- Setting rules limiting sources of contamination







Contamination control

Pre-PCR Post-PCR Physical barrier Air flow PCR boxes with UV lamps dUTP + uracil -N- glycosylase _ Psoralen + UV 10% caustic soda (surface decontamination) Tips with an aerosol barrier







Advantages of PCR

Speed (2-5 hours)

Simple, fast and cheap

Both DNA and RNA amplification

High amplification rate (10⁶ - 10⁹)

Theoretically 1 molecule

Highly specific

DNA can also be partially damaged (fragmentation)







The PCR product can be sequenced

The PCR product can be cloned into vectors

Amplification of fragments up to 30 kb long

Limitations of PCR

Highly susceptible to contamination and false positives

- **Complexity of primers**
- DNA sequence (Genebank)

Analysis of PCR products takes longer than the reaction itself







PCR modification

- **Nested PCR**
- **Multiplex PCR**
- Sequence-specific PCR
 - Allele specific
- Reverse-transcriptase PCR (RT-PCR) Long-range PCR
- Quantitative real-time PCR







Reverse transcription

Reverse transcription is the process of enzymatic conversion of an ssRNA molecule into complementary DNA (cDNA)







RT-PCR

Phases of the RT-PCR reaction :

- Isolation of RNA
- Reverse transcription
- PCR amplification
- PCR product analysis









Reverse transcription

Primers

- Oligo (dT)
- Random hexamers
- Random nonamers
- Classic primers

Reverse transcriptases

Retroviral RNA - dependent DNA polymerase

AMV Reverse transcriptase (Avian myeloblastosis virus)

MMLV reverse transcriptase (Moloney murine leukemia virus)

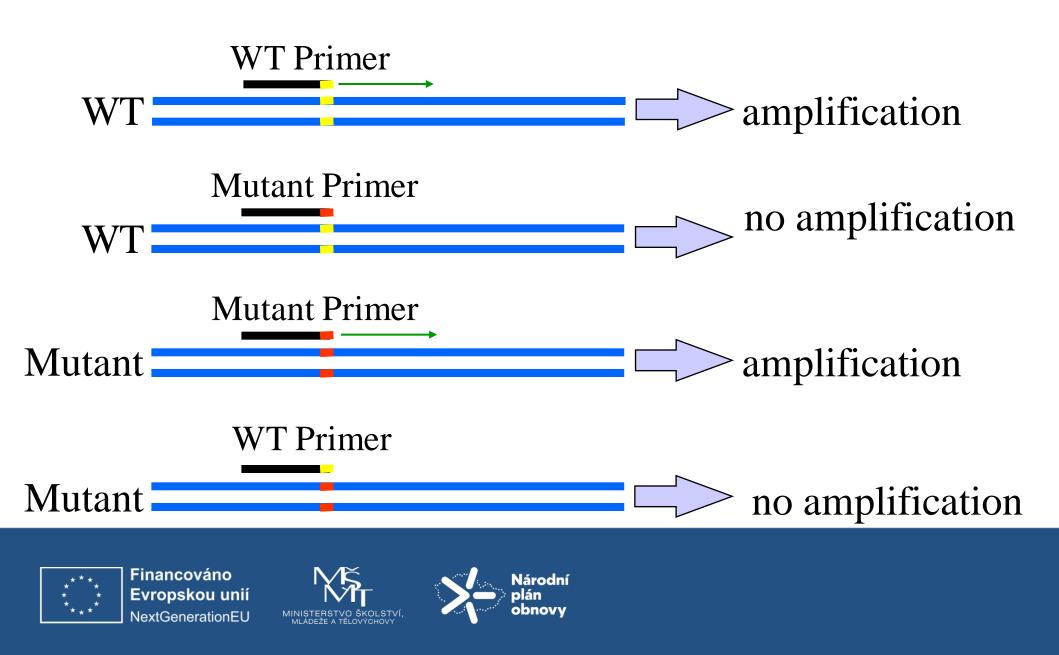




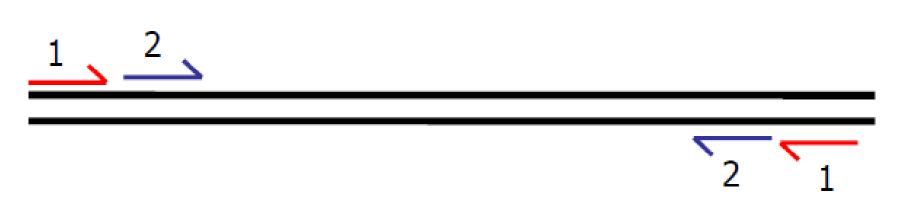




Allelic specific PCR amplification



Nested PCR



- A substantial gain in both specificity and sensitivity
- High risk of contamination







Multiplex PCR

- Simultaneous detection of multiple microorganisms (genes)
- Difficult optimization
- Medical diagnostics







REAL TIME PCR

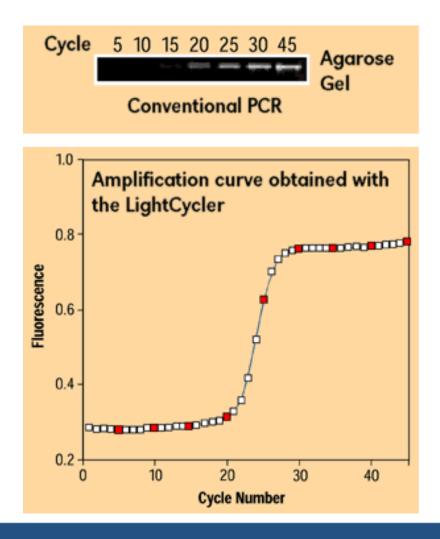
- Detection of PCR products
 - Intercalating dye (ethidium bromide, syber green dye): non-specific
 - Fluorogenic probes: more specific







Graph of dependence of fluorescence intensity on the number of PCR cycles







Quantitative PCR (qPCR)

The PCR product is exponentially multiplied (double in each cycle)

The PCR signal is recorded as an exponential curve described by phases: lag, log, linear and plateau

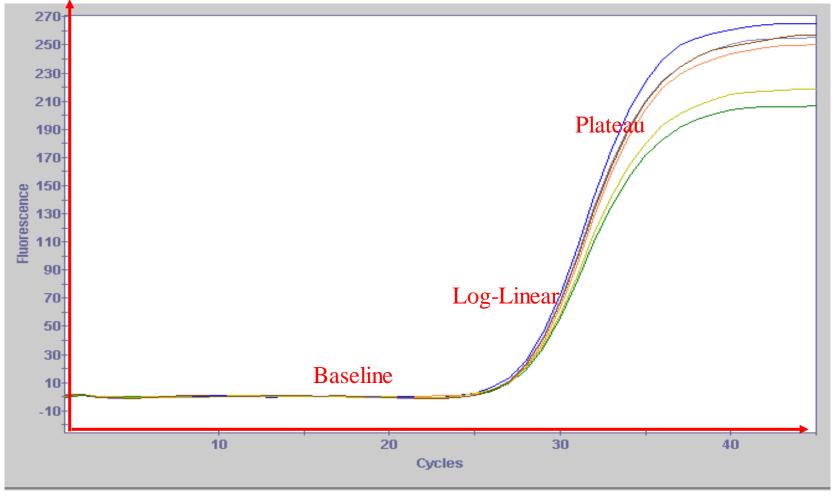
The length of the lag phase is inversely proportional to the amount of starting material.





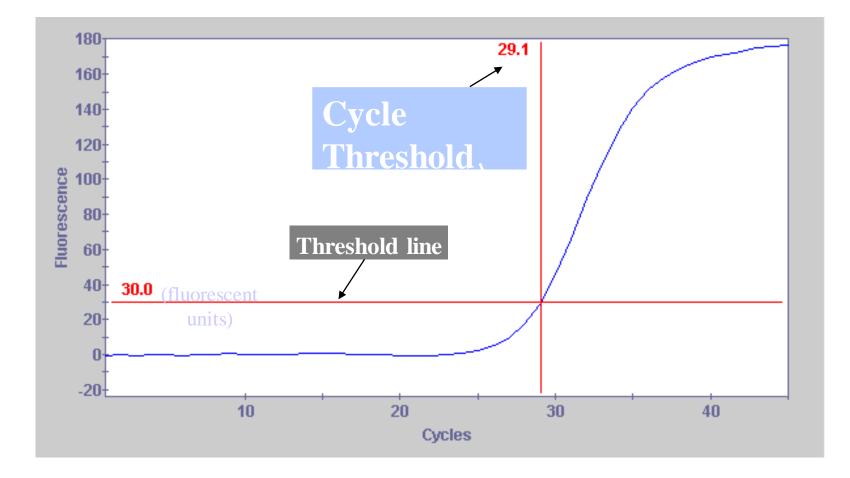


Progress of Real Time PCR





Threshold Cycle (Ct)

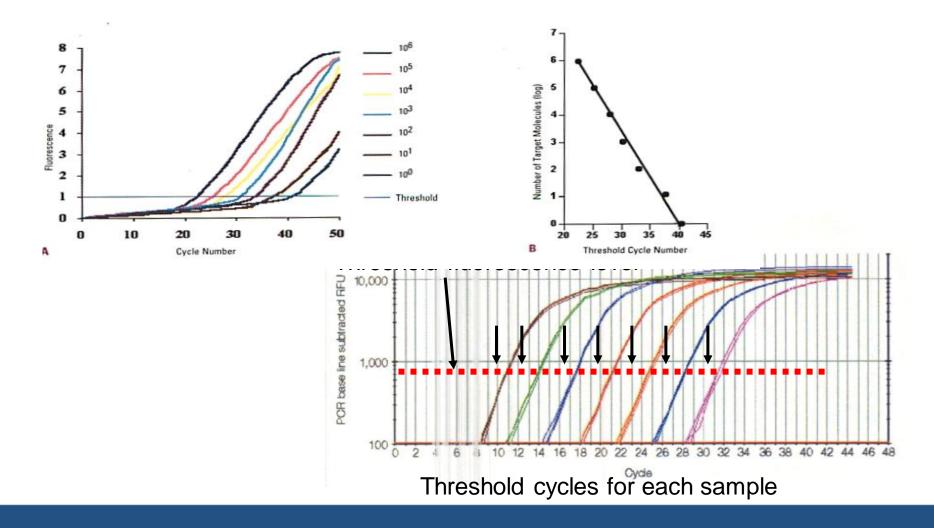








Construction of a standard curve







qPCR

DNA specific

- Ethidium bromide
- SyBr® green
- Hybridization probes
 - Hydrolyzing (TaqMan ®)
 - Molecular Beacons®, FRET®









Probes for Real-Time PCR

Hydrolysis probes

Taq Man probes

Fluorescent reporter at the 5' and quencher at the 3' end

Molecular beacons

Hair-like structure

Fluorescent reporter at the 5' and quencher at the 3' end

FRET probes

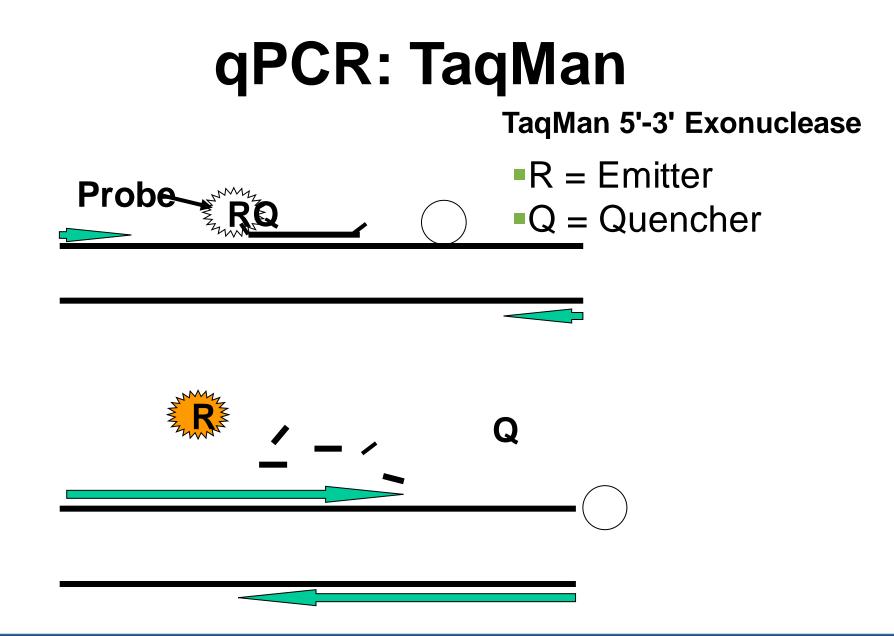
Fluorescence resonance energy transfer







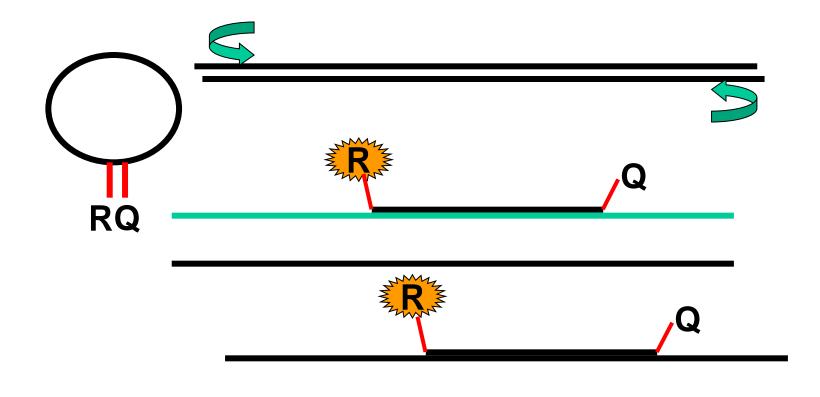








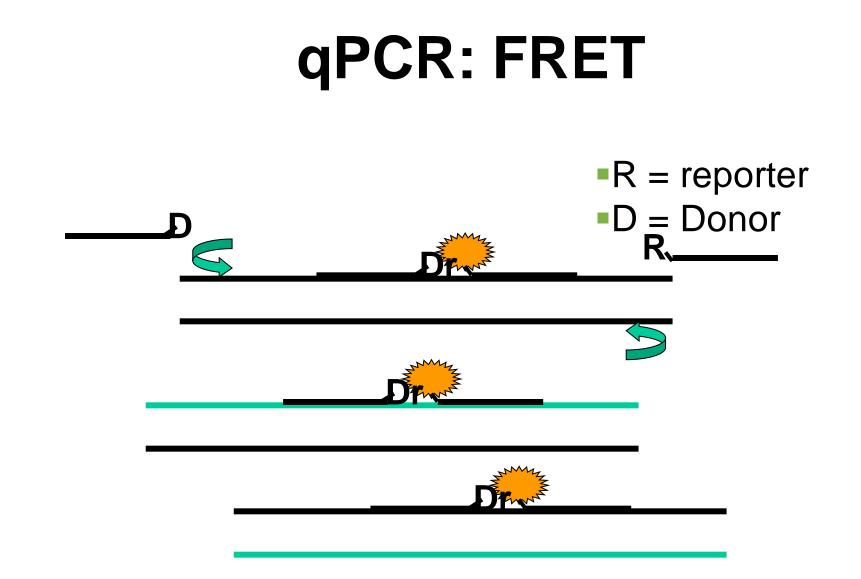
qPCR: Molecular Beacons







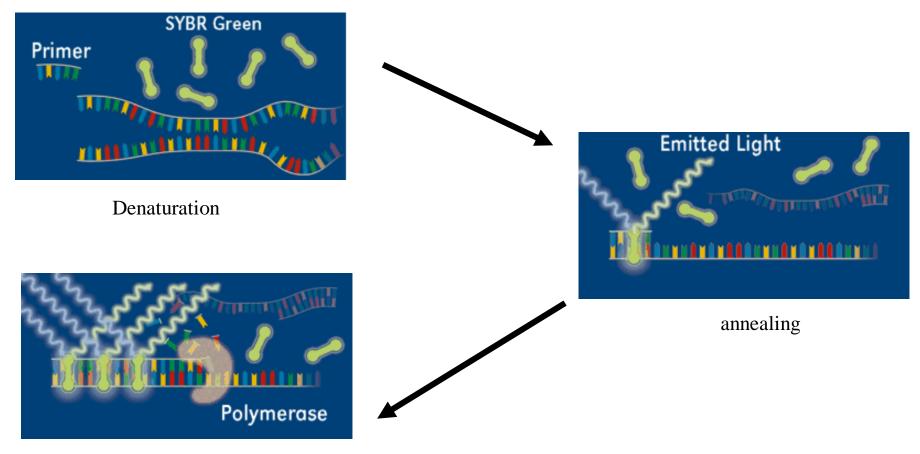








SYBR Green I



Národní

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Extension





qPCR systems

Cyclers with fluorescence detection and specialized software

The PCR reaction takes place in plates, test tubes or capillaries







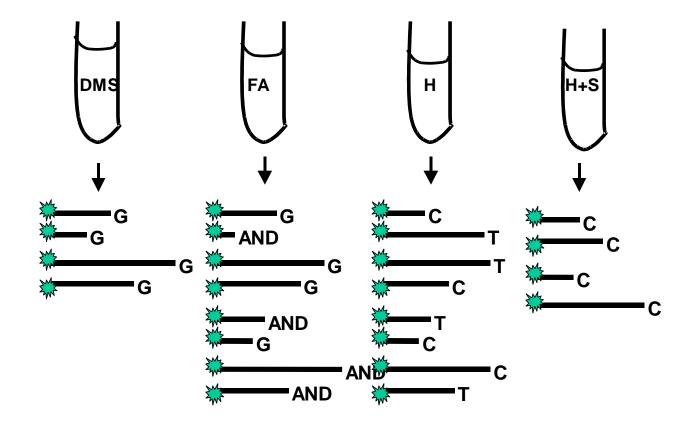
Sequencing







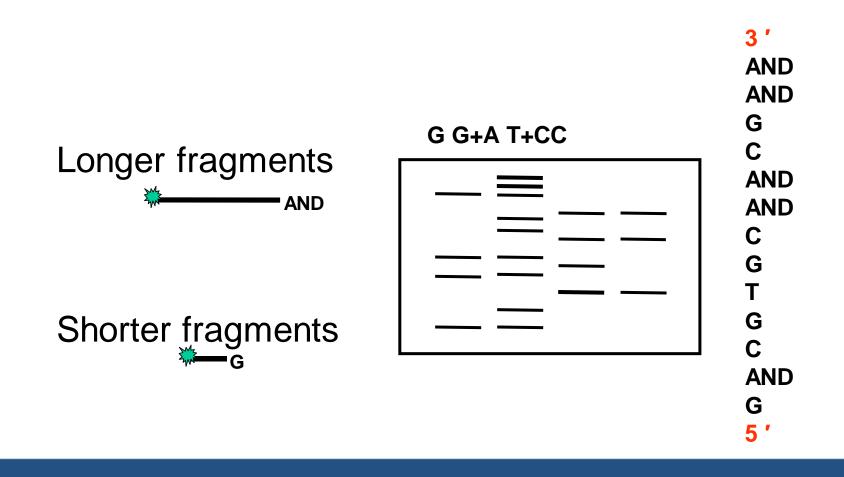
Maxam – Gilbert sequencing







Maxam - Gilbert









Chain Termination (Sanger) sequencing

A D	ddATP + four dNTPs	<mark>ddA</mark> dAdGdCdTdGdCdCdCdG
c	ddCTP + four dNTPs	dAdG <mark>ddC</mark> dAdGdCdTdG <mark>ddC</mark> dAdGdCdTdGdC <mark>ddC</mark> dAdGdCdTdGdCdC <mark>ddC</mark>
G	ddGTP + four dNTPs	dA <mark>ddG</mark> dAdGdCdT <mark>ddG</mark> dAdGdCdTdGdCdCdC <mark>ddG</mark>
т	<mark>ddTTP</mark> + four dNTPs	dAdGdC <mark>ddT</mark> dAdGdCdTdGdCdCdCdG





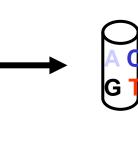


Dye Terminator Sequencing

Different fluorochromes are used

The reaction can thus be carried out in one test tube



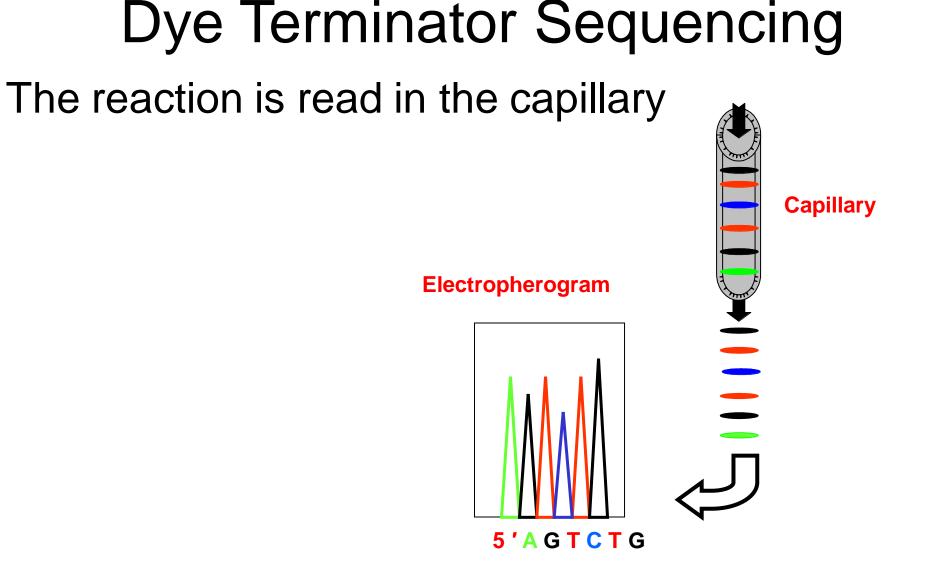


Fragments are distinguished based on size and "color"







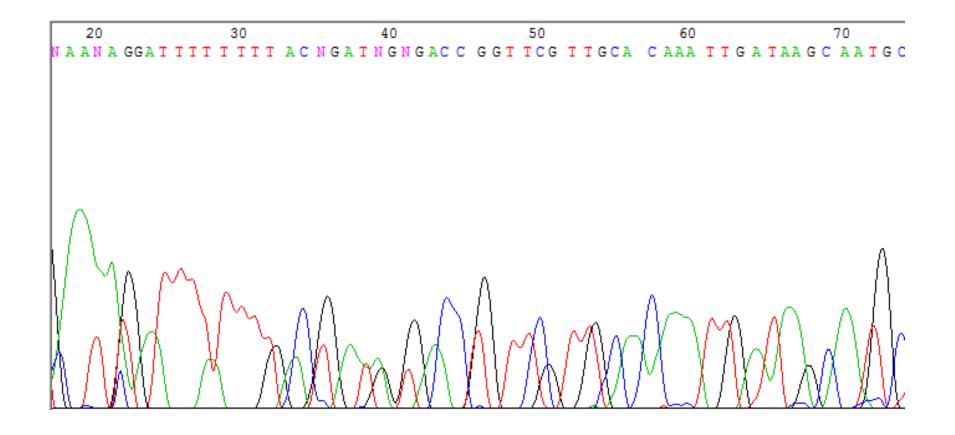














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RFLP (restriction fragment length polymorphism)









Restriction enzymes

Type I

Methylation / Cleavage (3 subunits) >1000 bp from the binding site eg Eco AI GAGNNNNNGTCA

Type II

Cleavage in specific locations

Type III

Methylation / Cleavage (2 subunits)

24–26 bp from the binding site

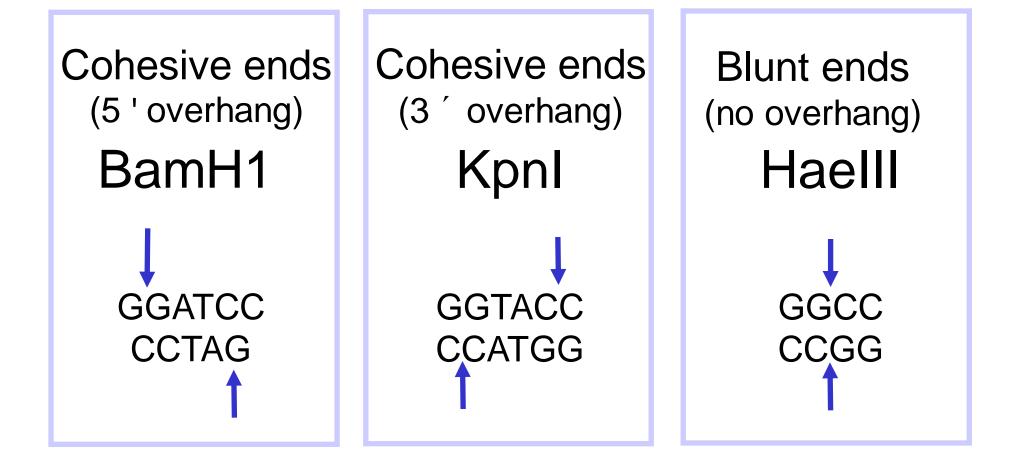
e.g. Hinf III CGAAT







Restriction endonucleases









Restriction enzyme mapping

- DNA cleavage by restriction endonucleases
- Electrophoresis
- The number of fragments indicates the number of restriction sites
- The size of the fragments indicates the distance between the cleavage sites

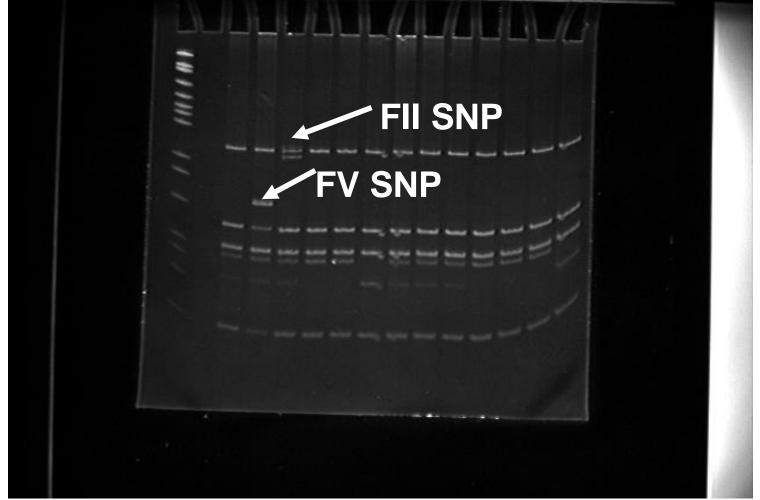








RFLP

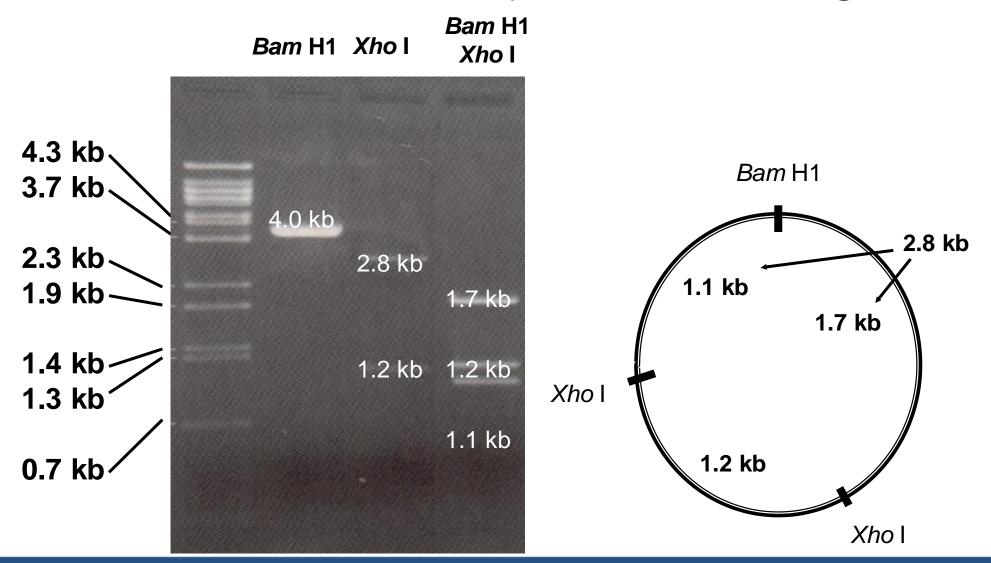








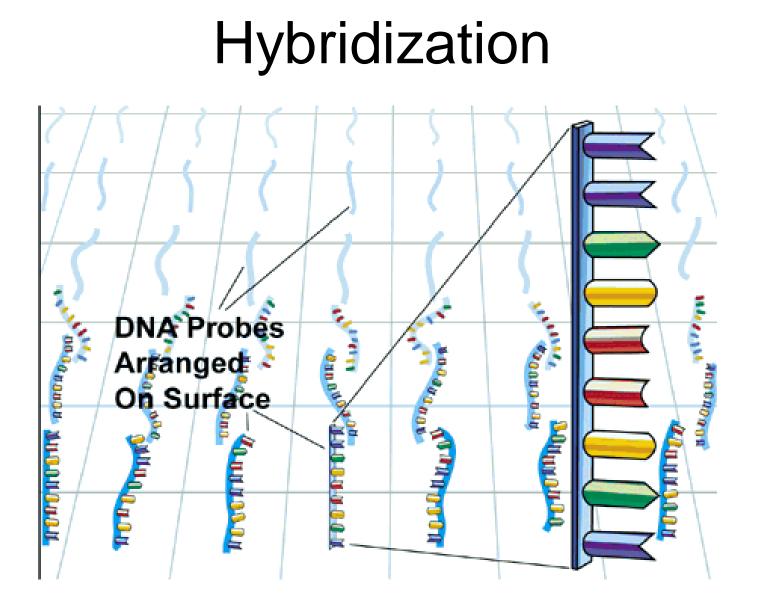
Restriction enzyme mapping









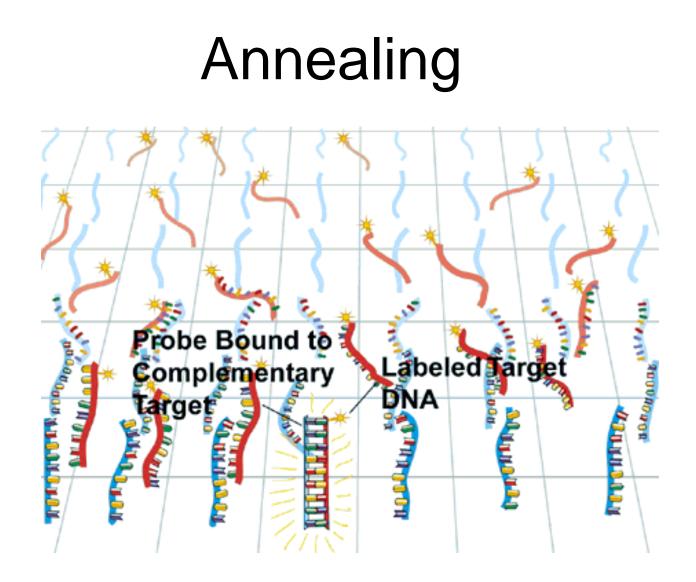










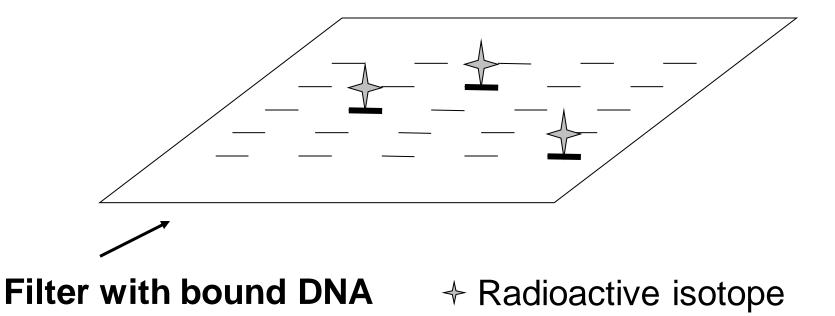








Radioactive detection



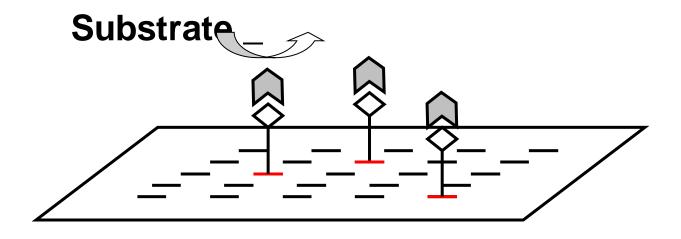
- Probe

novv





Non-radioactive detection

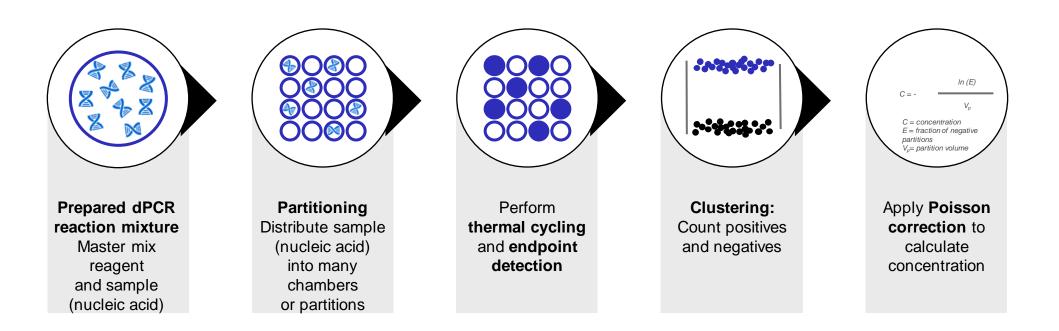


Antidigoxigenin antibody conjugated with kaline phosphatase or horseradish peroxidase Probe covalently labeled with biotin





digital PCR principle General workflow

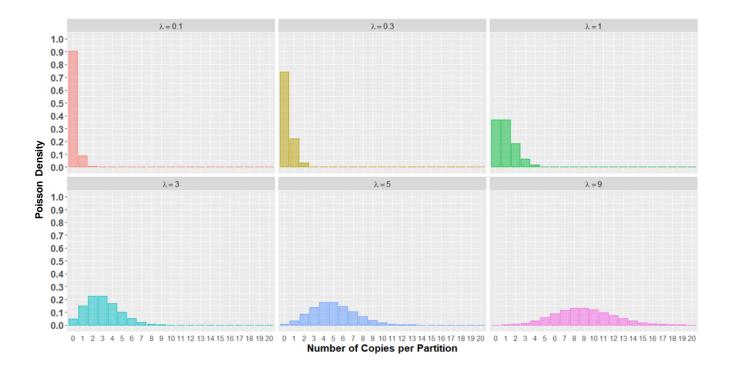








The dPCR Principle Poisson distribution

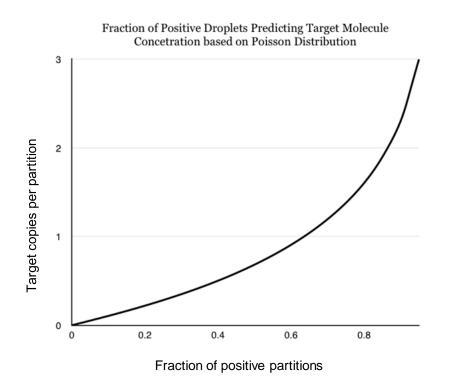


Poisson describes the probability that a partition contains k target molecules (k=0, 1, 2, ..., 20) for various target concentrations ($\lambda = 0.1, 0.3, 1, ...9$)





Poisson correction



 $\lambda = -\ln(1-P) = -\ln(E)$

- λ : Target copies/partition
- P: Fraction of positive partitions
- E: Fraction of negative partitions



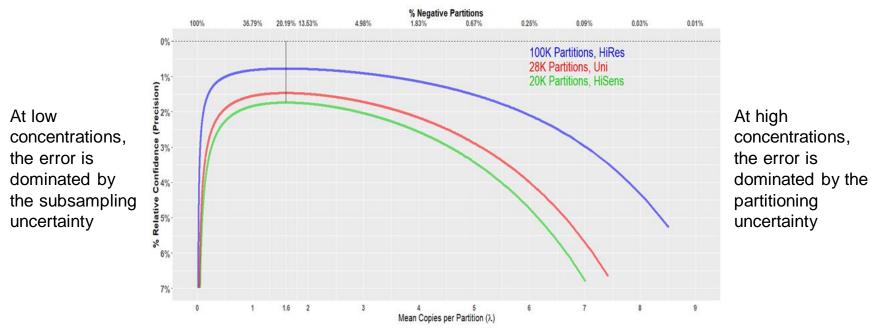
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Národní

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The dPCR Principle Quantification accuracy



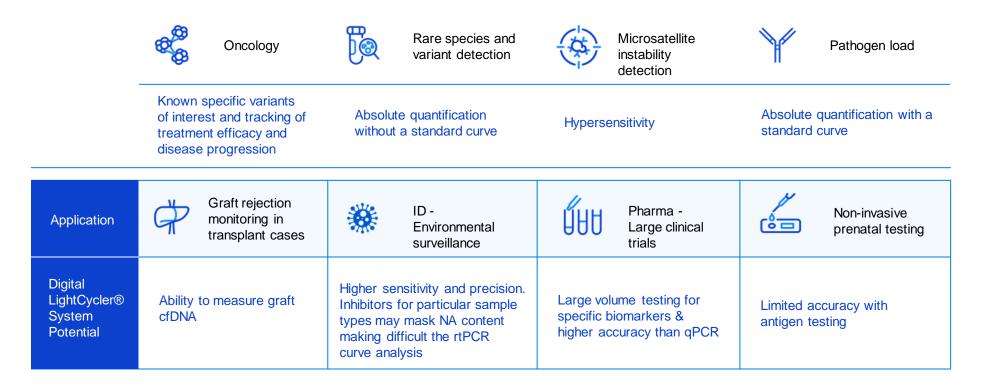
Quantification accuracy of dPCR

The accuracy of dPCR is <u>non-uniform</u> and depends on the average occupancy of the target sequence per partition (Lambda; λ). The accuracy of dPCR increases with an increasing number of partitions. The relative uncertainty is generally minimal at $\lambda \approx 1.6$ or 20% negative partitions and decreases with increasing partition numbers.





dPCR aplication Quantification accuracy









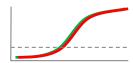


dPCR Advantages Accuracy and robustness

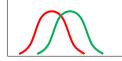
Scenario 1: Similar Number of Genome Copies



Real-time PCR 1,000 vs. 1,100 copies



Unable to distinguish differences in CT



Significant difference in Poisson distributions

Scenario 2:

Few Copies in Presence of Inhibitors



Few copies in presence of inhibitors



Inhibitors reduce amplification efficiency



Endpoint PCR where efficiency is not critical



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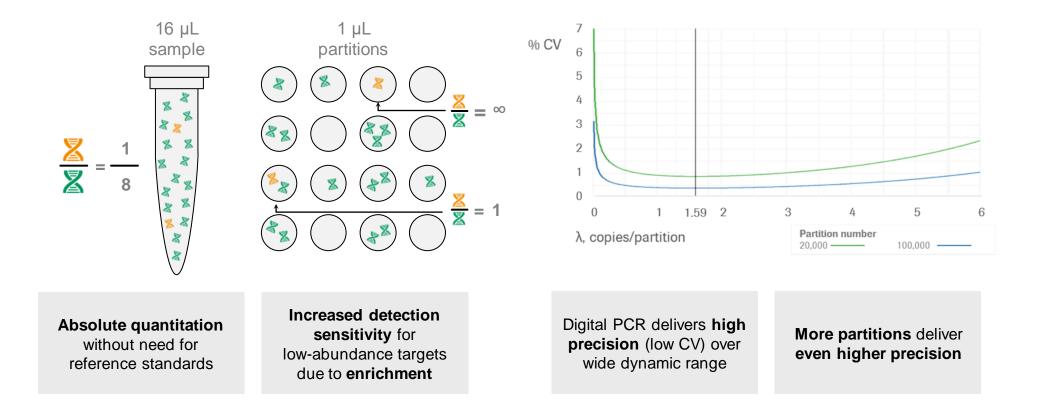




Quantitative PCR

dPCR

dPCR Advantages Reduced competition and high precision





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